

Delivery of antisense oligonucleotides using cholesterol-modified sense dendrimers and cationic lipids

Chalitin, Patrick; Margineanu, Anca; Marchand, Damien; Aerschot, Arthur Van; Rozenski, Jef; Schryver, Frans De; Hermann, Andreas; Müllen, Klaus; Juliano, Rudolph; Fisher, Michael H.

Published in:
BIOCONJUGATE CHEMISTRY

IMPORTANT NOTE: You are advised to consult the publisher's version (publisher's PDF) if you wish to cite from it. Please check the document version below.

Document Version

Publisher's PDF, also known as Version of record

Publication date:

2005

[Link to publication in University of Groningen/UMCG research database](#)

Citation for published version (APA):

Chalitin, P., Margineanu, A., Marchand, D., Aerschot, A. V., Rozenski, J., Schryver, F. D., Hermann, A., Müllen, K., Juliano, R., Fisher, M. H., Kang, H., Feyter, S. D., & Herdewijn, P. (2005). Delivery of antisense oligonucleotides using cholesterol-modified sense dendrimers and cationic lipids. *BIOCONJUGATE CHEMISTRY*, 16(4), 827 - 836.

Copyright

Other than for strictly personal use, it is not permitted to download or to forward/distribute the text or part of it without the consent of the author(s) and/or copyright holder(s), unless the work is under an open content license (like Creative Commons).

The publication may also be distributed here under the terms of Article 25fa of the Dutch Copyright Act, indicated by the "Taverne" license. More information can be found on the University of Groningen website: <https://www.rug.nl/library/open-access/self-archiving-pure/taverne-amendment>.

Take-down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Delivery of Antisense Oligonucleotides Using Cholesterol-Modified Sense Dendrimers and Cationic Lipids

Patrick Chaltin,[†] Anca Margineanu,[‡] Damien Marchand,[†] Arthur Van Aerschot,[†] Jef Rozenski,[†] Frans De Schryver,[‡] Andreas Herrmann,^{||} Klaus Müllen,^{||} Rudolph Juliano,[§] Michael H. Fisher,[§] Hyunmin Kang,[§] Steven De Feyter,[‡] and Piet Herdewijn^{†,*}

Laboratory for Medicinal Chemistry, Rega Institute, Catholic University of Leuven, Minderbroedersstraat 10, B-3000 Leuven, Belgium, Laboratory of Photochemistry and Spectroscopy, Catholic University of Leuven, Celestijnenlaan 200F, B-3000 Leuven, Belgium, Department of Pharmacology, University of North Carolina School of Medicine, 1106 Mary Ellen Jones Building Campus Box 7365, Chapel Hill, North Carolina 27599-7365, and Max-Planck-Institut für Polymerforschung, Ackermannweg 10, D-55021 Mainz, Germany. Received January 28, 2005; Revised Manuscript Received April 15, 2005

Cholesterol modified mono-, di-, and tetrameric oligonucleotides were synthesized and hybridized with antisense oligonucleotides to study their incorporation in cationic liposomes together with the influence of this dendrimeric delivery system on biological activity. Electrostatic interactions seem to play the most important role during complexation with cationic lipids. This oligonucleotide formulation gives a small but significant increase in the inhibition of P-glycoprotein expression in a cellular system.

INTRODUCTION

Antisense oligonucleotides are very powerful tools for inhibition of gene expression (1). The initial hurdles of enzymatic instability and poor hybridization strength of natural antisense oligonucleotides have been largely overcome by introducing chemical modifications. Delivery of oligonucleotides to target cells *in vivo*, however, is the major bottleneck for the widespread use of antisense oligonucleotides in therapy (2).

One of the potential solutions for this delivery problem is the use of liposomes (3). The antisense oligonucleotide (negatively charged due to the ionization of the phosphate backbone) interacts electrostatically with the cationic phospholipids forming the vesicles and is delivered to the cell by endocytosis. However, the number of oligonucleotides trapped into one complex is low. The delivery process can be improved by conjugation of the antisense oligonucleotide with cholesterol (4) or by the use of cholesterol for liposome formulations (5). A disadvantage of the first approach is that the cholesterol moiety is covalently attached to the oligonucleotide which influences the intracellular distribution of the antisense oligonucleotide.

Despite the efforts that have been made to characterize the formation of DNA-cationic liposome complexes (lipoplexes), the process is not yet completely understood. This is because multiple factors can influence their formation, such as the ratio between the number of negative charges in DNA and that of the positive charges in liposomes,

the fluidity of the lipid bilayer, and the formation of lipid domains after DNA binding (6). Furthermore, the size instability was correlated with high transfection efficiency *in vivo* (7). The current view about the organization of lipoplexes comes from electron microscopy and X-ray diffraction studies (8, 9). According to these, lipoplexes have a multilamellar structure with repeat distances equivalent to the thickness of a lipid bilayer and a single DNA double helix.

Here we investigated the approach of derivatization of an enzymatic labile sense–DNA with cholesterol and delivering the enzymatic stable antisense oligonucleotide as hybrid with this cholesterol–sense–DNA. As well as monomers, dimers and tetramers of sense–DNA were synthesized (Figure 1). Cationic lipids are used for the liposome formulation. This approach has the advantage that the enzymatic stable antisense oligonucleotide itself is not derivatized (i.e. its intracellular distribution and hybridization with its target is not influenced by a reporter group). The cholesterol-modified sense–DNA may stay locked in the endosomes and may be slowly degraded while the enzymatically more stable antisense oligonucleotide can diffuse into the cytoplasm and/or the nucleus (10). The synthesis was tested using an 11-mer as well as an 18-mer sense–DNA. For biological activity determination, we selected the 18-mer, as this length seems to be optimal for this purpose (10). The antisense oligonucleotide that is used as a model is a phosphorothioate 20 gapmer consisting of 2'-methoxyethyl (2'-MOE) nucleotides in the flanks and phosphorothioate DNA in the middle (5'-CCAUCCgacctcgCGCUCC-3'). The phosphorothioate DNA gap of eight nucleotides allows RNaseH to play a role in the mode of action of the antisense oligonucleotide. The phosphorothioate 2'-MOE flanks render the oligonucleotide stable against enzymatic degradation. This sequence has been demonstrated to inhibit P-glycoprotein expression in a cellular system (11).

* To whom correspondence should be addressed. Fax: +32 16 337341. Tel.: +32 16 337387. Piet.Herdewijn@rega.kuleuven.ac.be.

[†] Laboratory for Medicinal Chemistry, Rega Institute, Catholic University of Leuven.

[‡] Laboratory of Photochemistry and Spectroscopy, Catholic University of Leuven.

[§] University of North Carolina School of Medicine.

^{||} Max-Planck-Institut für Polymerforschung.

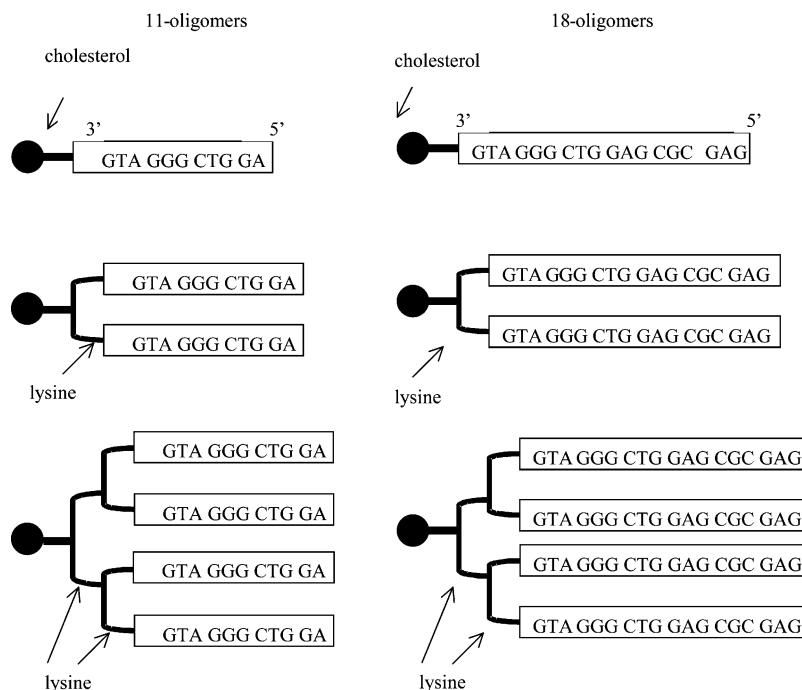


Figure 1. Schematic representation of monomeric and dendrimeric oligonucleotides (DNA) that were used to concentrate antisense oligonucleotides in cationic lipids for cellular delivery (as well, 11-mer as 18-mer oligonucleotides were evaluated as complexing agents for the antisense oligonucleotides).



Figure 2. Structure of antisense oligonucleotide directed at mRNA that codes for the P-glycoprotein.

EXPERIMENTAL PROCEDURES

Chemistry. ^1H and ^{13}C NMR spectra were recorded at 200 MHz with a Varian Gemini 200 in CDCl_3 . ^{13}C assignments were based on the usual shift increments and calculus values and where necessary confirmed by APT experiments. Assignments of quaternary carbons of the aromatic rings were based on comparison with other compounds and were mostly tentative. Electron impact mass spectra in positive (EI^+) and negative ion mode (EI^-) were acquired on a Kratos MS50TC mass spectrometer (Kratos, Manchester, UK), using MASPEC II data acquisition software (Mass Spectrometry Services Ltd., Manchester, UK). Electrospray ionization mass spectra in negative ion mode (ESI^-) were recorded on a quadrupole orthogonal acceleration/time-of-flight mass spectrometer (Q-ToF-2, Micromass, Manchester, UK). Absorption spectra were recorded with a Perkin-Elmer Lambda 40 spectrophotometer (Wellesley, MA) and fluorescence spectra with a Spex Fluorolog 3-22 fluorimeter (Jobin Yvon-Spex Instruments S. A., Inc., Edison, NJ). Precoated Machery-Nagel Alugram SilG/UV 254 plates were used for TLC, and spots were examined with UV light and a sulfuric acid/anisaldehyde spray. Column chromatography was performed on Acros silica gel (60-200 nm). Anhydrous solvents were obtained as follows: THF was refluxed on LiAlH_4 overnight and was distilled; dichloromethane (DCM) was stored on calcium hydride, refluxed, and distilled; pyridine, triethylamine (TEA), and *N,N*-diisopropylethylamine (DIEA) were refluxed overnight on potassium hydroxide and distilled. *N,N*-Dimethylformamide (DMF) was stored on activated molecular sieves (4 Å) for 3 days and was tested for absence of dimethylamine by the bromophenol test prior to use. DCM, DMF, acetic anhydride (Ac_2O), and pyridine were

obtained from BDH (Poole, England). Piperidine, trifluoroacetic acid (TFA), diisopropylcarbodiimide (DIC), 1-hydroxybenzotriazole (HOBt), 1-methylimidazole (NMI), di-*tert*-butyl dicarbonate [(Boc)₂O], benzyl chloroformate, 9-fluorenylmethyloxycarbonyl chloride (FmocCl), 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU), and *N*-methylmorpholine were supplied by ACROS (Geel, Belgium). 1-Hydroxy-7-azabenzotriazole (HOAt) and its tetramethyluronium derivative (HATU) were purchased from Perspective Biosystems. [2-(1*H*-benzotriazole-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate] (HBTU) from Advanced Chemtech. Benzotriazol-1-yloxytrispyrrolidinophosphonium hexafluorophosphate (PyBOP) was obtained from Advanced Chemtech. Diethyl azodicarboxylate (DEAD) and sodium hydride (NaH) were purchased from Sigma-Aldrich. *p*-[(*R,S*)- α -[1-(9*H*-fluoren-9-yl)methoxyformamido]-2,4-dimethoxybenzyl]phenoxyacetic acid (Fmoc-Rink linker) was supplied by Novabiochem. Lipofectamine 2000 was obtained from Invitrogen. PBS buffer (pH7.4) was obtained from Sigma. Oligonucleotide assembly was performed on an Expedite DNA synthesizer (Applied Biosystems) using phosphoramidites from Glen Research.

The 11 and the 18 oligonucleotide sense sequences without cholesterol, as well as the tetramethyl rhodamine (TMR)-labeled oligonucleotide antisense sequence, complementary to the gene encoding the P-glycoprotein (11), were obtained from Invitrogen. The phosphorothioate oligonucleotide from Figure 2 is a gift from ISIS.

Cholesterylacetaldehyde Diethylacetal (2). To a solution of cholesterol (5 g, 12.9 mmol, 1 equiv) in a mixture of anhydrous THF/DMF (1:1, 200 mL) were added NaH (3.2 g, 77.6 mmol, 6 equiv) and, after 5 min, bromoacetaldehyde diethylacetal (6.3 mL, 38.8 mmol, 3

equiv) under anhydrous conditions. A nitrogen-flushed reflux condenser was attached, and the mixture was heated at reflux for 18 h. The excess NaH was decomposed using MeOH/CH₂Cl₂ and mild shaking. After evaporation to dryness, silica gel-based chromatographic purification was performed with a stepwise gradient of a hexane/EtOAc mixture (98:2 to 95:5) as eluent, yielding 4.3 g (8.56 mmol, 66.3%) of the title compound. ¹³C NMR: δ = 140.97 (C-3), 121.58 (C-4), 101.46 (β -CH), 79.73 (C-1), 68.77 (α -CH₂), 62.09 (Et, CH₂O), 56.72 (C-7), 56.12 (C-10), 50.11 (C-14), 42.25 (C-11), 39.73; 39.45; 38.97 (C-2; C-12; C-24), 37.15 (C-16), 36.78 (C-15), 36.12 (C-22), 35.72 (C-20), 31.84 (C-5, C-6), 28.26; 28.13; 27.92 (C-17; C-9; C-25), 24.19; 23.73 (C-8; C-23), 22.73; 22.46 (C-26; C-27), 20.97 (C-13), 19.27 (C-18), 18.60 (C-21), 15.27 (Et, CH₃), 11.75 (C-19). Mass calcd for [M + Cl]⁻ C₃₃H₅₈O₃Cl₁: 537.4074, found: (ESI⁻): 537.4094.

Cholesterylacetaldehyde (3). To a solution of **2** (4.1 g, 8.17 mmol) in CH₂Cl₂ (80 mL) was added 20 mL of TFA/H₂O (1:1), and the reaction mixture was stirred for 4.5 h at room temperature (complete conversion). NaOH (1 N) was used to neutralize the solution, and subsequently an extraction with CH₂Cl₂ was performed. The organic layer was dried over anhydrous MgSO₄, filtered, and evaporated in vacuo to dryness, yielding 3.45 g (8.07 mmol, 98%) of the title compound. ¹³C NMR: δ = 201.70 (β -CH), 140.30 (C-3), 122.22 (C-4), 80.37 (C-1), 73.81 (α -CH₂), 56.69 (C-7), 56.12 (C-10), 50.08 (C-14), 42.25 (C-11), 39.70; 39.42; 38.76 (C-2; C-12; C-24), 36.97 (C-16), 36.72 (C-15), 36.12 (C-22), 35.69 (C-20), 31.81 (C-5, C-6), 28.10; 27.92 (C-17; C-9; C-25), 24.19; 23.73 (C-8; C-23), 22.70; 22.46 (C-26; C-27), 20.97 (C-13), 19.24 (C-18), 18.60 (C-21), 11.75 (C-19). Mass calcd for [M]⁺ C₂₉H₄₈O₂: 428.3654, found (EI⁺): 428.3647.

Cholesterylacetic Acid (4). Compound **3** (3 g, 7.0 mmol) was dissolved in acetone (105 mL), and 1.5 mL Jones reagent (26.72 g CrO₃/23 mL concentrated H₂SO₄/H₂O to 100 mL) was added dropwise while the reaction mixture was cooled on ice. If necessary (TLC), more Jones reagent was added to completely convert **3** into its carboxylic acid. After 10 min, acetone and MeOH were added, the mixture was filtered, and the remaining filtrate was evaporated to dryness. The residual solid was purified by column chromatography on silica gel by elution with hexane/EtOAc/AcOH (8:2:0.1), yielding 2.1 g (4.76 mmol; 68%) of the title compound. ¹³C NMR: δ = 173.54 (COOH), 140.00 (C-3), 122.46 (C-4), 80.46 (C-1), 65.16 (α -CH₂), 56.69 (C-7), 56.12 (C-10), 50.05 (C-14), 42.25 (C-11), 39.67; 39.45; 38.61 (C-2; C-12; C-24), 36.90 (C-16), 36.69 (C-15), 36.12 (C-22), 35.69 (C-20), 31.78 (C-5, C-6), 28.13; 28.01; 27.92 (C-17; C-9; C-25), 24.19; 23.73 (C-8; C-23), 22.73; 22.46 (C-26; C-27), 20.97 (C-13), 19.24 (C-18), 18.60 (C-21), 11.75 (C-19). Mass calcd for [M]⁺ C₂₉H₄₈O₃: 444.3603, found (EI⁻): 444.3606.

N α -Fmoc-N ϵ -cholesterylacetyl-lysine-O-Me (5). Compound **4** (1.7 g, 3.7 mmol, 1 equiv) and HATU (1.35 g, 4.07 mmol, 1.1 equiv) were suspended in 85 mL of dry DMF. To this suspension was added DIEA (1.3 mL, 7.4 mmol, 2 equiv), and the resulting solution was stirred for 2 h at room temperature. To this mixture was added a solution of Fmoc-Lysine-OMe.HCl (1.7 g, 4.07 mmol, 1.1 equiv) and DIEA (1.3 mL, 7.4 mmol, 2 equiv) in 15 mL of dry DMF, and stirring was continued at room temperature for 18 h. After evaporation, the residue was dissolved in EtOAc, and the organic phase was washed with HCl (1 N), water, saturated Na₂CO₃ solution, brine, and water. The organic layer was dried over sodium sulfate, filtered, and evaporated. The remaining oil was purified by chromatography using a mixture of CH₂Cl₂/

MeOH (99:1) as eluent. Evaporation of the pooled fractions yielded 2.5 g (3.1 mmol, 84%) of compound **5**. ¹³C NMR: δ = 172.66 (Lys, COOMe), 170.41 (CONH); 156.51 (OCONH), 144.04 (Fmoc, C-10/11), 141.73 (Fmoc, C-12/13), 140.12 (Chol, C-3), 127.68 (Fmoc, C-3/6), 127.04 (Fmoc, C-2/7), 125.07 (Fmoc, C-1/8), 122.28 (Chol, C-4), 119.97 (Fmoc, C-4/5), 80.39 (Chol, C-1), 67.47 (Fmoc, CH₂O), 66.52 (Chol, α -CH₂), 56.66 (Chol, C-7), 56.08 (Chol, C-10), 52.38 (Lys, CH₃O), 51.14 (Lys, α -CH) 50.01 (Chol, C-14), 47.22 (Fmoc, C-9), 42.24 (Chol, C-11), 40.61 (Lys, ϵ -CH₂NH), 39.67; 39.45; 38.82 (Chol, C-2; C-12; C-24), 36.93 (Chol, C-16), 36.69 (Chol, C-15), 36.12 (Chol, C-22), 35.69 (Chol, C-20), 32.26 (β -CH₂(Lys)), 31.80 (Chol, C-5, C-6), 29.62 (δ -CH₂(Lys)), 28.13; 28.01; 27.92 (Chol, C-17; C-9; C-25), 24.19; 23.73 (Chol, C-8; C-23), 22.73; 22.46 (Chol, C-26; C-27), 22.34 (γ -CH₂(Lys)) 20.97 (Chol, C-13), 19.27 (Chol, C-18), 18.63 (Chol, C-21), 11.74 (Chol, C-19). Mass calcd for [M + Cl]⁻ C₅₁H₇₂N₂O₆Cl₁: 843.5078, found: (ESI⁻): 843.5049.

N α -Fmoc-N ϵ -cholesterylacetyl-lysine (6). The cleavage of the methyl ester was carried as described in the literature (12). To compound **5** (1.5 g, 1.8 mmol, 1 equiv) in 20 mL of THF at 0 °C was added 0.2 N LiOH (18 mL, 3.6 mmol, 2 equiv), and the reaction was stirred for 40 min. To the reaction mixture, 50 mL of a biphasic of 0.2 N HCl/EtOAc (1:1) was added, aqueous layer was extracted three times with EtOAc, and the combined organic layers were dried over MgSO₄, filtered, and evaporated to dryness. The remaining product was purified by column chromatography on silica gel with a CH₂Cl₂/MeOH mixture (95:5) as eluent, yielding 1.1 g (1.4 mmol, 77%) of the title compound. ¹³C NMR: δ = 174.78 (Lys, COOH), 171.14 (CONH); 156.51 (OCONH), 144.01 (Fmoc, C-10/11), 141.37 (Fmoc, C-12/13), 140.06 (Chol, C-3), 127.71 (Fmoc, C-3/6), 127.07 (Fmoc, C-2/7), 125.10 (Fmoc, C-1/8), 122.31 (Chol, C-4), 120.00 (Fmoc, C-4/5), 80.46 (Chol, C-1), 67.31 (Fmoc, CH₂O), 66.52 (Chol, α -CH₂), 56.63 (Chol, C-7), 56.08 (Chol, C-10), 51.47 (Lys, α -CH) 49.98 (Chol, C-14), 47.19 (Fmoc, C-9), 42.24 (Chol, C-11), 40.58 (Lys, ϵ -CH₂NH), 39.67; 39.45; 38.82 (Chol, C-2; C-12; C-24), 36.90 (Chol, C-16), 36.69 (Chol, C-15), 36.12 (Chol, C-22), 35.72 (Chol, C-20), 32.26 (β -CH₂(Lys)), 31.80 (Chol, C-5, C-6), 29.62, 29.19 (δ -CH₂(Lys)), 28.16; 28.01; 27.92 (Chol, C-17; C-9; C-25), 24.19; 23.76 (Chol, C-8; C-23), 22.73; 22.46 (Chol, C-26; C-27), 22.34 (γ -CH₂(Lys)) 20.97 (Chol, C-13), 19.24 (Chol, C-18), 18.63 (Chol, C-21), 11.74 (Chol, C-19). Mass calcd for [M-H]⁻ C₅₀H₆₉N₂O₆: 793.5155, found: (ESI⁻): 793.5143.

Solid-Phase Synthesis. Peptide Synthesis. Synthesis of the conjugates was performed on the LCAA-CPG carrier. For each amino acid or compound to be coupled, different reaction conditions were applied. After every reaction, the solution was filtered and the beads were washed with DMF (two times), CH₂Cl₂ (2 times), and acetonitrile (two times). The beads were dried, and Fmoc- or MMTr- substitution levels were determined. Hereto, an accurately measured quantity of resin was suspended in exactly 25 mL of 20% piperidine in DMF or in a HClO₄/EtOH mixture (3:2), respectively. After 15 min the absorbance was measured at 300 nm for fulvene (or adduct) determination and at 475 nm for methoxytrityl cation detection (ϵ_{Fmoc} = 7500, ϵ_{MMTr} = 55000). If the coupling yields were unsatisfactory, a second coupling reaction was performed. Before each new coupling, the remaining amino groups were capped using 1.5 mL of 20% acetic anhydride in THF and 1.5 mL of 20% 1-methylimidazole and 40% 2,4,6-trimethylpyridine in THF for 15 min, and washed with acetonitrile and DMF. Fmoc-deprotection was carried out with a piperidine/

DMF mixture (2:8 v:v, 2 mL) for 5 min and washing for 15 min, while for MMTr-deprotection, 3% TCA in CH_2Cl_2 was used 3 to 4 times for 3 min, until no yellow color was detectable.

Coupling of Fmoc-sarcosine to LCAA-CPG was generally performed as described in the literature (13). To a solution of Fmoc-sarcosine (116 mg, 0.37 mmol, 4 equiv), HOBt (31 mg, 2.23 mmol, 2.5 equiv), and HBTU (88 mg, 0.23 mmol, 2.5 equiv) in 2 mL DMF was added DIEA (78 μL , 0.46 mmol, 5 equiv), and the mixture was shaken for 5 min. More DIEA (78 μL , 0.46 mmol, 5 equiv) was added, and the solution was then poured onto LCAA-CPG beads (1 g, 92 μmol , 1 equiv). The reaction mixture was occasionally shaken for 2 h. Coupling of MMTr-butyric acid was obtained by adding a solution of 4-OMMTr-butyric acid (14) (220 mg, 0.46 mmol, 5 equiv), HOBt/HBTU (88 mg, 0.23 mmol, 2.5 equiv), and DIEA (158 μL , 0.92 mmol, 10 equiv) in 2 mL of DMF. Following deprotection of MMTr, Fmoc-glycine was coupled to the 4-OH group of 4-OH-butyric acid by occasionally swirling the beads with a reaction mixture of Fmoc-glycine (273 mg, 0.92 mmol, 10 equiv), DIC (72 μL , 0.46 mmol, 5 equiv), and DMAP (17 mg, 13.8 μmol , 0.15 equiv) in DMF (2 mL).

The cholesterylacetamide **6** was attached to glycine by using a solution of **6** (0.29 mg, 0.37 mmol, 4 equiv), HBTU (140 mg, 0.37 mmol, 4 equiv), HOBt (49 mg, 0.37 mmol, 4 equiv), and DIEA (168 μL , 0.92 mmol, 10 equiv) in 1 mL of DMF. The mixture was shaken for 5 min before it was added to the Fmoc-deprotected support (yield: 90%).

For single oligonucleotide loading, 4-O-MMTr-butyric acid was coupled as a linker before starting oligonucleotide synthesis. For the branching-off purposes, N α -Fmoc-N ϵ -Fmoc-lysine (4 equiv) was used. The protected lysine was attached to the support by adding a solution of HBTU (4 equiv), HOBt (4 equiv), and DIEA (10 equiv) in DMF. Subsequently, 4-O-MMTr-butyric acid was coupled. The amount of 4-O-MMTr-butyric acid used was always 4 times the amount of free NH_2 groups available for coupling.

Oligonucleotide Synthesis. Four different supports were used for assembly of the undeca- and octadecamers. The number of hydroxyl groups per gram of support was determined by detritylation and amounted to 48, 80, and 130 $\mu\text{mol/g}$ for the supports **9**, **11**, **13**, respectively. The assembly of oligonucleotides was done on the Expedite following a standard 1 μmol protocol, using only 0.5 μmol of support and a 4 min coupling time. Following deprotection with AMA (methylamine–ammonia 1:1) for 2 h at 25 °C, the products were purified by gel filtration using 30% aqueous acetonitrile to avoid adsorption of the cholesteryl moieties to the support. Depending whether a single, double, or four chains of oligonucleotides needed to be coupled to the cholesterol moiety, one of the different supports **9**, **11**, **13** were used. The conjugates could be purified by RP-HPLC on PLRP–S (100 Å, 8 μM , 250 \times 7 mm, Polymer Laboratories) with a CH_3CN gradient in 0.05 M ammonium acetate, pH 7.0. In general, in ascending order, tetrameric oligo-containing dendrimer eluted before the trimer, dimer, and monomer species. Analysis was accomplished by HPLC/MS on a capillary chromatograph (CapLC, Waters, Milford, CA). Columns of 150 mm \times 0.3 mm length (LCPackings, San Francisco, CA) were used. Oligonucleotides were eluted with a triethylammonium-1,1,1,3,3,3-hexafluoro-2-propanol:acetonitrile solvent system, and the flow rate was 5 $\mu\text{L/min}$. Electrospray spectra were acquired on a quadrupole orthogonal acceleration/time-of-flight mass spectrometer (Q-Tof-2, Micromass, Manchester, U.K.) in

Table 1. Results of Mass Spectrometry Analysis of the Monomeric (10), Dimeric (12), and Tetrameric (14) Constructs

	a series		b series	
	calcd	found ^c	calcd	found ^c
monomer	6433.4 ^a	6432.8	4226.1 ^a	4225.8
dimer	12358.4 ^b	12356.4	7937.8 ^a	7937.2
tetramer	24202.6 ^b	ND ^d	15368.9 ^b	15367.0

^a Monoisotopic mass. ^b Average mass. ^c Mass obtained after deconvolution of the electrospray spectrum. ^d ND: not determined.

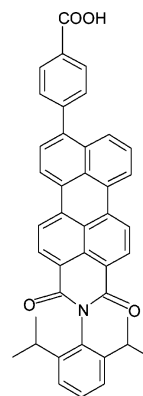


Figure 3. Structure of the perylene monoimide derivative used as a membrane probe.

negative ion mode. The scan time used was 2 s. The combined spectra from a chromatographic peak were deconvoluted using the MaxEnt algorithm of the software Masslynx 3.4 (Micromass). Theoretical oligonucleotide masses were calculated using the monoisotopic element masses for masses up to 8000 and using the average element masses for masses above 8000. The data are given in Table 1.

Fluorescence Resonance Energy Transfer (FRET) Measurements. Lipofectamine 2000 was diluted in PBS (phosphate buffer saline, pH = 7.4) (Sigma) and was labeled with a perylene monoimide derivative (PMI-COOH) (Figure 3) in an approximate ratio of 1 fluorescent molecule for 250 lipid molecules. Hybridization between the antisense oligonucleotide labeled with TMR and the DNA with or without cholesterol was performed by heating appropriate amounts in PBS at 80° for 3 min, followed by cooling at room temperature for 15 min. Hybridized DNA was added in increasing concentrations to the Lipofectamine suspension. The fluorescence energy transfer between PMI-COOH and TMR was monitored by exciting the sample at 450 nm (where the direct excitation of TMR is minimal) and measuring the fluorescence quenching of PMI-COOH at the maximum emission peak (549 nm). The values were corrected for the nonlabeled DNA quenching of PMI-COOH fluorescence and for the TMR emission at 549 nm.

Cells. NIH 3T3 cells stably transfected with a plasmid containing the human *mdr1* gene (pSK1 MDR) were gift from M. M. Gottesman. The MDR-3T3 cells were grown in DMEM (Dulbecco minimal cututred media) medium containing 10% FBS (fetal bovine serum) and 60 mg/mL colchicine.

Treatment of Cells with Antisense Oligonucleotides. The experimental protocols were similar to those previously described (11). Briefly, MDR 3T3 cells were grown in 162 mm flasks to 95% confluency and then seeded onto six-well plates at 3×10^5 /well in 10% FBS/DMEM and incubated 24 h. The cells were washed twice with PBS. Complexes between dendrimers and antisense

strand were prepared in 10 mM TRIS, 50 mM NaCl, 1 mM EDTA by heating the solutions at 80 °C and cooling to room temperature for 15 min. The complexation of the double stranded dendrimeric form of the antisense was done with Lipofectamine 2000 (Invitrogen) in OptiMEM according to standard procedure. The complexes were added to cells in 10% FBS/DMEM. The cells were incubated with complexes for 4 h. Then the cells were rinsed and further incubated for 68 h in 2% FBS/DMEM. All conditions were performed in triplicate. The P-glycoprotein percent shift is calculated from the lipofectamine 2000 control shift.

Pharmacological Effects of Antisense Treatment.

Analysis of the pharmacological effects of dendrimeric delivery was based on a previously described assay (15) involving antisense inhibition of P-glycoprotein expression in mouse 3T3 cells stably transfected with the human *MDR1* gene. The cell surface expression of P-glycoprotein was determined by immunostaining and quantitation by flow cytometry, as described (16). Briefly, the MRK16 anti-P-glycoprotein antibody (Kamiya Biochemicals), which is directed against an external epitope, was employed as the primary antibody. After the incubation, the cells were washed twice in 10% FBS/PBS. The level of phycoerythrin (PE)-conjugated goat anti-mouse IgG (Sigma, St. Louis, MO) was used as the second antibody in viable cells (viability determined by light scatter) was quantitated using the Summit V3.0 software application (Cytomation Inc.) on a Becton Dickinson flow cytometer.

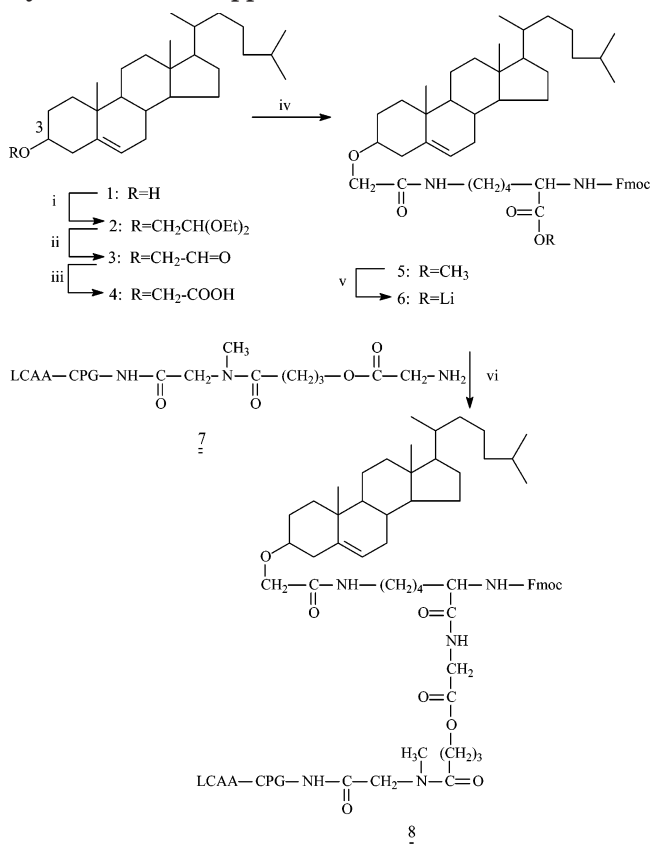
RESULTS AND DISCUSSION

Synthesis. For the synthesis of the monomeric, dimeric, and tetrameric cholesterol oligonucleotide conjugates, a strategy was used based on lysine chemistry. Lysine has an α - and ϵ -amino group, and binding of one lysine unit via its α carboxyl function to a solid support allows the two functional amino groups to be used for oligonucleotide synthesis.

Cholesterol (**1**) was derivatized at its C-3 hydroxyl group with a functional group that is stable against acid and base and that can be used for the formation of an amide bond (Scheme 1). Reaction of cholesterol with chloroacetic acid in the presence of NaH gives poor yields. Therefore, cholesterol was first reacted with the diethylacetal of bromoacetaldehyde in the presence of NaH in a mixture of THF/DMF for 18 h at 85 °C. In this way cholesterylacetaldehyde diethylacetal (**2**) was obtained in 70% yield. Reaction of **2** with Jones reagent resulted in the formation of the desired cholesterylacetic acid **4**, but the reaction went to completion only after 24 h, and multiple side products were formed. To avoid side reactions, the acetal was first hydrolyzed with trifluoroacetic acid in dichloromethane for 4 h, giving the aldehyde **3**. The oxidation of the aldehyde to the carboxylic acid with Jones reagent was completed after 10 min, giving a clean reaction product. The oxidation using hypochlorite was less effective.

Subsequently, Fmoc-Lysine-OMe was coupled with its free ϵ -NH₂-group to cholesteryl acetic acid using HATU, DIEA, and TEA (Scheme 1). Compound **5** was then deprotected by hydrolyzing the methyl ester with 0.2 N LiOH in THF/H₂O. The carboxylic acid of **6** could then be used to couple **6** to the solid support **7**. This solid support material was obtained by derivatization of LCAA-CPG with sarcosine. Subsequently, MMTr-O-butyric acid was attached to the sarcosine and detritylated

Scheme 1. Synthesis of Cholesterol-Conjugated Lysine on Solid Support^a



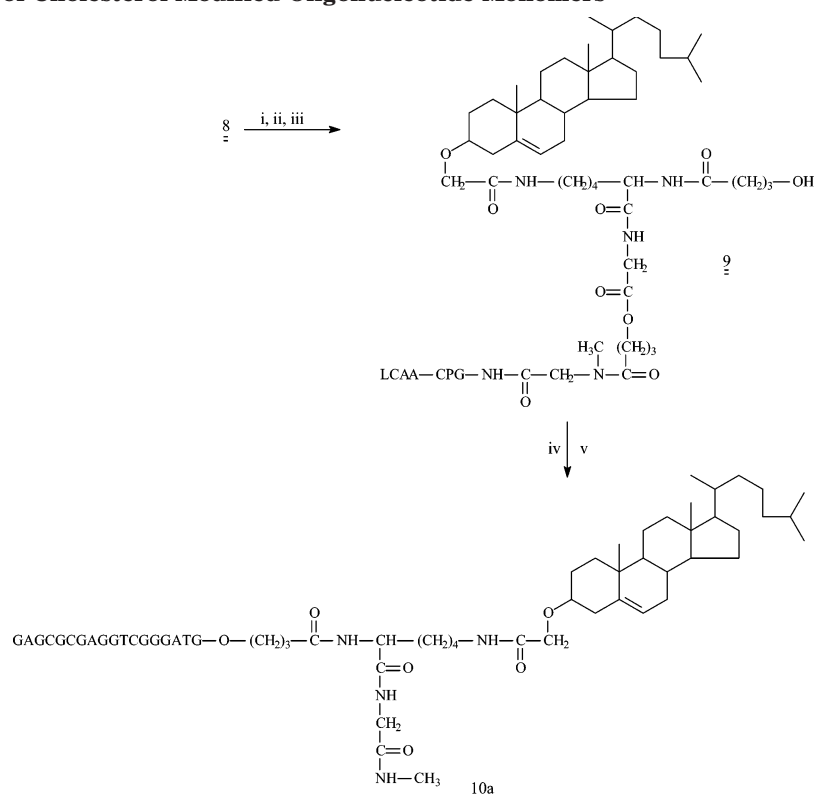
^a i: BrCH₂CH(OEt)₂, NaH, DMF/THF; ii: CF₃COOH aqueous in DCM; iii: CrO₃, H₂SO₄, CH₃COCH₃, H₂O; iv: Fmoc-Lysine-OMe HCL, HATU, DIEA, DMF; v: LiOH, THF, H₂O; vi: HBTU, HOBT, DIEA, DMF.

with trifluoroacetic acid. The free hydroxyl group was used for coupling with Fmoc-glycine with formation of a base-labile ester bond. After Fmoc deprotection, compound **6** was coupled to the solid support **7** using a mixture of HBTU, HOBT, and DIEA (Scheme 1). In this way, a cholesterol-derivatized lysine connected to a solid support via an ester bond was obtained, as general support for the synthesis of the desired oligonucleotide monomers (**10a,b**) and dendrimers (**12a,b** and **14a,b**).

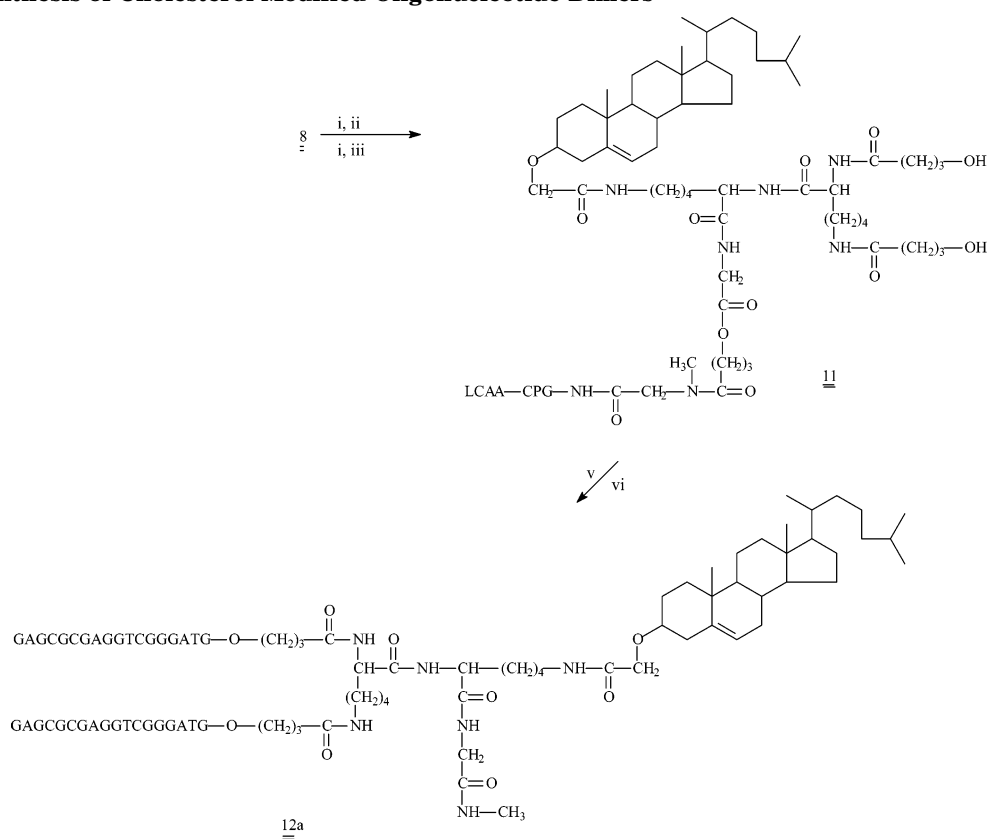
The synthesis of the cholesteryl-lysyl oligonucleotide (monomer **10a,b**) is shown in Scheme 2. After Fmoc deprotection of **8**, coupling of MMTr-O-butyric acid, and MMTr deprotection, the solid material **8** was used for oligonucleotide synthesis with the phosphoramidite approach. An 11-mer and 18-mer were assembled with a sequence complementary to the antisense oligonucleotide.

Deprotection using AMA reagent (concentrated aq ammonia-aq methylamine 1:1) yielded **10a,b**. The conjugate was concomitantly removed from the solid support by cleavage of the labile ester group with formation of the methylamide (with 5% amide formation according to MS analysis).

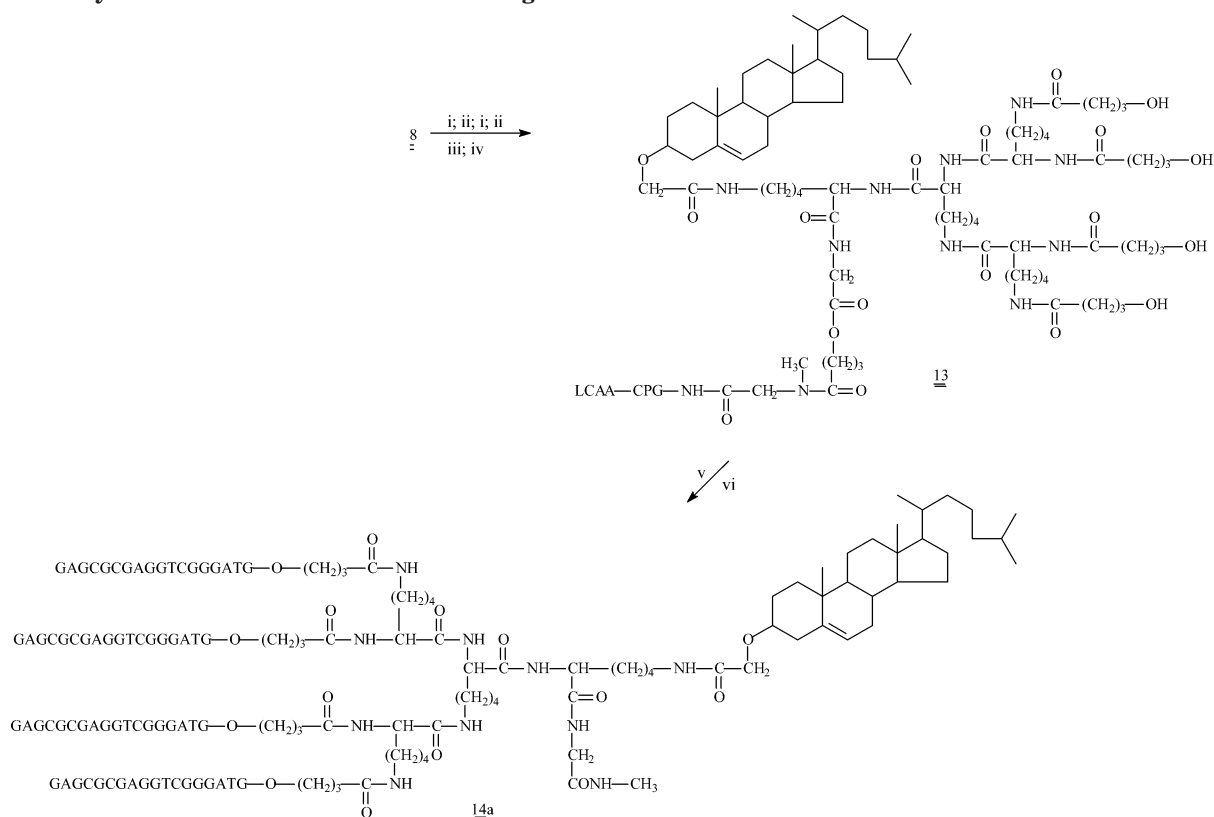
Branching of the peptide linker was carried out by coupling of Fmoc-protected lys-OH on deprotected **8** (Scheme 3). After a second deprotection step and binding of hydroxybutyric acid, the solid support can be used for the synthesis of two oligonucleotides on the same cholesteryl derivatized peptide (Scheme 3). Repetition of the protocol of coupling Fmoc-protected lys-OH, Fmoc deprotection, and coupling of hydroxybutyric acid might lead to an exponential branching of the cholesterol-derivatized

Scheme 2. Synthesis of Cholesterol-Modified Oligonucleotide Monomers^a

^a i: piperidine, DMF; ii: MMTro(CH₂)₃COOH, HOBT, HBTU, DIEA, DMF; iii: CCl₃COOH, DCM; iv: oligonucleotide synthesis; v: deprotection (aq. CH₃NH₂-NH₃) **10b**: same molecule with oligo sequence: AGGTCGGGATG-.

Scheme 3. Synthesis of Cholesterol-Modified Oligonucleotide Dimers^a

^a i: piperidine, DMF; ii: (Fmoc)₂lys-OH, HBTU, HOBT, DIEA, DMF; iii: MMTro(CH₂)₃COOH, HOBT, HBTU, DIEA, DMF; iv: CCl₃COOH, DCM; v: oligonucleotide synthesis; vi: deprotection (aq. CH₃NH₂-NH₃) **12b**: same construct with oligo sequence: AGGTCGGGATG-.

Scheme 4. Synthesis of Cholesterol-Modified Oligonucleotide Tetramers^a

^a i: piperidine, DMF; ii: (Fmoc)₂lys-OH, HBTU, HOBT, DIEA, DMF; iii: MMTrO(CH₂)₃COOH, HOBT/HBTU, DIEA, DMF; iv: CCl₃COOH, DCM; v: oligonucleotide synthesis; vi: deprotection procedure (aq. CH₃NH₂-NH₃). **14b**: same construct with oligo sequence: AGGTCGGGATG.

solid support, enabling the assembly of multiple oligonucleotides on one cholesterol moiety. This is demonstrated with the branched dendrimer **14a,b** in Scheme 4. However, mass spectral analysis of the major fraction shows that only three of the four hydroxybutyryl moieties are used for oligonucleotide synthesis. The HPLC sample of the trimeric compound shows only a small fraction of the four-branched dendrimer. The three-branched molecule has an acetylated lysine moiety (although it is not clear if acetylation has occurred at the α -N or the ϵ -N of lysine). Apparently, steric hindrance or the occurrence of a side reaction hampers the formation of the four-branched construct. We were able, however, to isolate sufficient amounts of the tetrameric construct by chromatographic purification to carry out biological tests.

Using these three different supports, cholesterol oligonucleotide complexes were formed in a ratio of 1:1 (**10a,b**), 1:2 (**12a,b**), and 1:4 (**14a,b**), and the obtained products were used for hybridization with the antisense oligonucleotide followed by cationic liposome complexation. Although it is difficult to analyze the way the antisense oligonucleotides are associated to the sense-dendrimers, the stability of the complex was determined using melting experiments. In all cases, the melting point was above 70 °C, demonstrating that stable complexes are formed.

Lipoplexes Formation Followed by FRET. The dynamics of DNA-liposomes interaction can be monitored in a FRET experiment due to the sixth power dependence of the fluorescence energy transfer efficiency to the distance between donor and acceptor. The process can thus take place on distances of a few nanometers, meaning that only the DNA bound to the surface of liposomes will contribute to the variation of the measured

signal. The FRET approach was used to investigate the association between DNA and liposomes with different compositions (17), to follow in time the changes in lipid-DNA interaction during lipoplex formation (18), or to monitor the oligonucleotide release into living cells (19). Theoretical calculations of FRET efficiency particularly applied for the geometry of lipoplexes were also developed (20).

In the context of our ongoing study of perylene monoimide derivatives in biological systems, this type of fluorescent molecule has been used in these experiments for several reasons. Due to its hydrophobic rylene core, the molecule is rapidly incorporated into a lipid bilayer, while the hydrophilic carboxylic group most probably anchors the molecule at the phospholipid-water interface. This dye was chosen instead of other fluorescent probes for membranes (e.g. diphenyl hexatriene DPH or its derivatives) because of the very high fluorescence quantum yield (almost 1) and the high photostability in hydrophobic environments of the PMI chromophore (21), making possible the detection of fluorescence intensity variations at very low concentrations (10⁻⁸–10⁻⁷ M). The molecule is practically devoid of fluorescence in water, so the dye that was not incorporated into liposomes does not contribute to the total intensity measured (ongoing study). In addition, the carboxylic group is negatively charged at the pH used for these experiments (pH 7.4), which makes the interaction with the positively charged lipids composing the Lipofectamine more efficient when compared to an indocarbocyanine dye with two long alkyl chains (DiI-C18) (data not shown). Third, the high spectral overlap between the PMI-COOH emission and TMR absorption (Figure 4) makes this dye pair very good for FRET measurements.

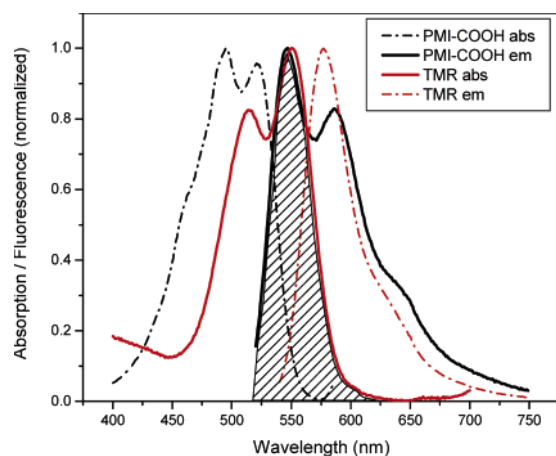


Figure 4. Absorption and emission spectra for PMI-COOH and TMR. The filled area represents the overlap between the emission spectrum of PMI-COOH and the absorption spectrum of TMR. The overlap integral calculated according to the Förster relation (22) is $J = 5.2 \times 10^{-13} \text{ cm}^6 \text{ mmol}^{-1}$ and the Förster radius is $\approx 60 \text{ Å}$.

The purpose of our experiments was to observe if cationic liposomes (Lipofectamine) are able to complex more antisense DNA hybridized with the cholesterol-modified sense sequence than the same DNA hybridized with the normal sense oligonucleotide (10). Lipofectamine suspension labeled with the donor (PMI-COOH) was kept at constant concentration in all experiments, while increasing acceptor concentrations (hybridized TMR-DNA) were added to the sample. Figure 5a and b show the averages and the standard deviations obtained from three independent experiments for each double strand type [naked DNA (11 DNA and 18 DNA) and cholesterol-modified monomeric (10a,b), dimeric (12a,b), and tetrameric (14a,b) DNA were evaluated]. The intensity of the donor in the presence of the acceptor (I_{DA}) was normalized to the initial intensity of the donor (I_D). Using a logarithmic scale, the first data points in each determination could be linearly fitted, and the values obtained for the slopes are given in Table 2.

As can be observed, hybridizing the 20-mer sequence with the cholesterol-modified sense 11-mer oligonucleotides (10b, 12b, 14b) shifts the slopes of the linear fitting toward higher values when compared to the unmodified sense sequence. Assuming that each TMR molecule acts as an independent acceptor even when two (12b) or four (14b) DNA oligonucleotides are linked in the same

Table 2. Slopes for the Linear Fit of Data in Figure 5

DNA sequence	slope	DNA sequence	slope
11DNA	$(5.14 \pm 0.33) \times 10^6$	18DNA	$(5.68 \pm 0.54) \times 10^6$
10b	$(6.37 \pm 0.35) \times 10^6$	10a	$(5.85 \pm 0.30) \times 10^6$
12b	$(5.70 \pm 0.20) \times 10^6$	12a	$(5.94 \pm 0.42) \times 10^6$
14b	$(6.15 \pm 0.47) \times 10^6$	14a	$(5.82 \pm 0.43) \times 10^6$

dendrimer, more donor molecules are quenched for the same DNA amount added to the system when cholesterol-modified sequences are used. Thus, the antisense oligonucleotide quantity bound to the liposomes is slightly higher in the case of cholesterol-modified sense oligonucleotides. The differences between the unmodified and the cholesterol-modified sequence are not very pronounced, but they are statistically significant. On the other hand, it seems that the number of oligonucleotides linked to the same cholesterol molecule does not influence the binding. When the 18-mer sense sequence is used for hybridization, no differences in the uptake in lipoplexes can be observed between the cholesterol-modified (10a, 12a, 14a) and the unmodified oligonucleotides. Another observation is that the slope values obtained for the 11- and 18-oligonucleotide sequence are not statistically different. Given the small length change from the 11 to the 18 sequences, we can conclude that the electrostatic interaction is the most important factor governing the formation of lipoplexes, while the much weaker hydrophobic interaction due to the presence of cholesterol may have a role in the case of short DNA sequences.

The fact that we were not able to find significant differences in the liposome loading does not necessarily mean that a higher transfection efficiency in living cells cannot be obtained with the cholesterol-modified sequences. Previous studies reported a higher transfection efficiency obtained with cholesterol-modified oligonucleotides by themselves due to an increased binding to the cellular membranes (23, 24). It seems thus possible that the uptake and the metabolism pathways, even when using a delivery agent, are different for the cholesterol-modified sequences when compared to the usual ones.

Biological Activity. The biological activity was determined using the 2'-MOE gapmer of phosphorothioate as the antisense strand (Table 3). The concentration was determined spectroscopically. When incubating the antisense oligonucleotide with Lipofectamine 2000, a maximum activity was obtained at 50 nM (giving 42.2% P-gp shift). Therefore, this antisense concentration was used in further experiments. When the cholesterol dendrimers

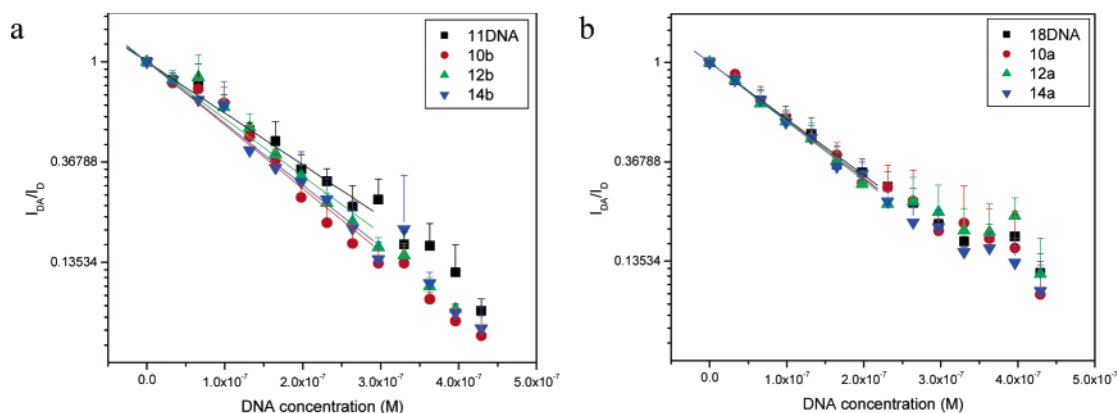


Figure 5. Quenching of PMI-COOH fluorescence (labeling Lipofectamine) using the 11 (a) and the 18 (b) oligonucleotide sequence for hybridization of the TMR-antigen sequence. The intensity of the donor in the presence of the acceptor (I_{DA}) was normalized to the initial intensity of the donor (I_D). Note that logarithmic scale was used. The first data points in each determination could be linearly fitted, and the values obtained for the slopes are given in Table 2.

Table 3. Percent Reduction of P-Glycoprotein Expression Level in Mouse 3T3 Cells by Dendrimer–Antisense Complexes^a

	antisense alone	dimer 12a + antisense ratio 1:1	tetramer 14a + antisense ratio 1:1	dimer 12a + antisense ratio 1:2	tetramer 14a + antisense ratio 1:4
P-gp	62.8%	68.9%	70.5%	72.4%	74.0%

^a NIH3T3-MDR cells were transfected with the complexes of **12a** or **14a**, the antisense oligonucleotide, and Lipofectamine 2000. Results were obtained after 48 h incubation in 5% FBS. The ratio between **12a** or **14a** and the antisense oligonucleotide was 1:1, 1:2, and 1:4. The antisense oligonucleotide was always used at 50 nM concentration.

12a and **14a** were tested in complex with the antisense construct but without adding lipofectamine 2000, no significant effect was seen except after very long incubation time (72 h). This means that the dendrimers alone were not effective in delivering the antisense oligonucleotide. Significant effects were obtained when the dendrimers **12a** and **14a** were used in the presence of lipofectamine 2000 (Table 3). After 48 h incubation the antisense oligonucleotide gives 62.8% P-gp shift, while in the presence of the dendrimers and using 1:1, 1:2, or 1:4 molar ratios of dendrimer and antisense oligonucleotides (fixed at 50 nM), the P-gp shift varied from 68.9% to 74.0%. The effect is modest but significant. The biological activity is somewhat increased when the antisense oligonucleotides are more condensed (ratio 1:1 versus ratio 1:2 or 1:4).

CONCLUSION

Using a linker based on lysine chemistry, we were able to synthesize monomeric, dimeric, and tetrameric oligonucleotide constructs covalently linked to a cholesterol moiety. These monomeric and dendrimeric sense DNA were complexed with enzymatic stable antisense oligonucleotides targeting P-glycoprotein expression. Perylene dye was used to monitor uptake of the constructs in cationic lipids. In the case of 11-mer cholesterol-modified sense DNA, a higher amount of antisense oligonucleotide could be incorporated in the cationic liposomes than without cholesterol modification. However, this difference did not exist with the 18-mer cholesterol-modified sense DNA. Also the number of oligonucleotides linked to the same cholesterol moiety does not influence binding to the cationic lipids. In this case, electrostatic interactions seems to play the most important role during complex formations and longer sense DNA gives higher concentration of antisense DNA in the lipoplexes. The previously observed results of the better delivery of duplex oligonucleotide versus single-stranded oligonucleotides (10) may be partially explained by this phenomenon. However, dendrimer delivery of the antisense oligonucleotide using cholesterol-modified 18-mer sense DNA and lipofectamine resulted in a small but significant increase in biological activity. This increase in biological activity is, most probably, due to a difference in intracellular metabolism i.e., the kinetics with which the antisense oligonucleotide is delivered to its target mRNA.

ACKNOWLEDGMENT

We thank René De Boer for assistance with the mass spectrometric analysis, and Chantal Biernaux and Tom De Roeck for their excellent editorial help. Steven De Feyter is a postdoctoral fellow of the Fund for Scientific Research – Flanders. The Catholic University of Leuven (IDO/02/014) is acknowledged for financial support. Financial support from the University of North Carolina was provided by the grant P01GM59259 to R. L. Juliano. The authors thank the Federal Science Policy through IUAP-V-03, and the Institute for the promotion of innovation by Science and Technology in Flanders (IWT).

Anca Margineanu gratefully acknowledges the Research Foundation of KU Leuven through GOA 2001/2.

LITERATURE CITED

- (1) Scherer, L. J., and Rossi, J. J. (2003) Approaches for the sequence-specific knockdown of mRNA. *Nat. Biotechnol.* **21**, 1457–1465.
- (2) Shoji, Y., and Nakashima, H. (2004) Current status of delivery systems to improve target efficacy of oligonucleotides. *Curr. Pharm. Des.* **10**, 785–796.
- (3) Boulikas, T. (1996) Liposome DNA delivery and uptake by cells. *Oncol. Rep.* **3**, 989–995.
- (4) Bijsterbosch, M. K., Manoharan, M., Dorland, R., Waarlo, I. H., Biessen, E. A., and Van Berkel, T.-J. (2001) Delivery of cholesteryl-conjugated phosphorothioate oligodeoxynucleotides to Kupffer cells by lactosylated low-density lipoprotein. *Biochem. Pharmacol.* **62**, 627–633.
- (5) Girão de Cruz, M. T., Simões, S., Pedrosa de Lima, M. C. (2004) Improving lipoplex-mediated gene transfer into C6 glioma cells and primary neurons. *Exp. Neurol.* **187**, 65–75.
- (6) Zuhorn, I. S., and Hoekstra, D. J. (2002) On the mechanism of cationic amphiphile-mediated transfection. To fuse or not to fuse: is that the question? *J. Membr. Biol.* **189**, 167–179.
- (7) Barenholz, Y. (2001) Liposome application: problems and prospects. *Curr. Opin. Colloid Interface Sci.* **6**, 66–77.
- (8) Simberg, D., Danino, D., Talmon, Y., Minsky, A., Ferrari, E., Wheeler, C. J., and Barenholz, Y. (2001) Phase behavior, DNA ordering, and size instability of cationic lipoplexes. Relevance to optimal transfection activity. *J. Biol. Chem.* **276**, 47453–47459.
- (9) Radler, J. O., Koltover, I., Salditt, T., and Safinya, C. R. (1997) Structure of DNA-cationic liposome complexes: DNA intercalation in multilamellar membranes in distinct interhelical packing regimes. *Science* **275**, 810–814.
- (10) Astriab-Fisher, A., Fisher, M. H., Juliano, R., and Herdewijn, P. (2004) Increased uptake of antisense oligonucleotides by delivery as double stranded complexes. *Biochem. Pharmacol.* **68**, 403–407.
- (11) Astriab-Fisher, A., Sergueev, D. S., Fisher, M., Shax, B. R., and Juliano, R. L. (2000) Antisense inhibition of P-glycoprotein expression using peptide–oligonucleotide conjugates. *Biochem. Pharmacol.* **60**, 83–90.
- (12) Qabar, M. N., Urban, J., and Kahn, M. (1997) A facile solution and solid-phase synthesis of phosphotyrosine mimetic L-4-[diethylphosphono(difluoromethyl)]-phenylalanine (F(2)-Pmp(Eto)(2)) derivatives. *Tetrahedron* **53**, 11171–11178.
- (13) Stetsenko, D. A., and Gait, M. (2001) A convenient solid-phase method for synthesis of 3'-conjugates of oligonucleotides. *Bioconjugate Chem.* **12**, 576–586.
- (14) Hovinen, J., Guzaev, A., Azharyev, A., and Lonnberg, H. (1994) Novel solid supports for the preparation of 3'-derivatized oligonucleotides: Introduction of 3'-alkylphosphate tether groups bearing amino, carboxy, carboxamido and mercapto functionalities. *Tetrahedron* **50**, 7203–7218.
- (15) Alahari, S. K., Dean, N. M., Fisher, M. H., Delong, R., Manoharan, M., Trivel, K. L., and Juliano, R. L. (1996) Inhibition of expression of the multidrug resistance-associated P-glycoprotein of by phosphorothioate and 5' cholesterol-conjugated phosphorothioate antisense oligonucleotides. *Mol. Pharmacol.* **50**, 508–519.
- (16) Alahari, S. K., Delong, R., Fisher, M. H., Dean, N. M., Viliet, P., and Juliano, R. L. (1998) Novel chemically modified oligonucleotides provide potent inhibition of P-glycoprotein expression. *J. Pharmacol. Exp. Ther.* **286**, 419–428.

- (17) Wiethoff, C. M., Gill, M. L., Koe, J. G., and Middaugh, C. R. (2002) The structural organization of cationic lipid-DNA complexes. *J. Biol. Chem.* 277, 44980–44987.
- (18) Zhang, Y., Garzon-Rodriguez, W., Manning, M. C., and Anchordoquy, T. J. (2003) The use of fluorescence resonance energy transfer to monitor dynamic changes of lipid-DNA interactions during lipoplex formation. *Biochim. Biophys. Acta* 1614, 182–192.
- (19) Zelphati, O., and Szoka, F. C., Jr. (1996) Mechanism of oligonucleotide release from cationic liposomes. *Proc. Natl. Acad. Sci. U.S.A.* 93, 11493–11498.
- (20) Madeira, C., Loura, L. M. S., Aires-Barros, M. R., Fedorov, A., and Prieto, M. (2003) Characterization of DNA/lipid complexes by fluorescence resonance energy transfer. *Biophys. J.* 85, 3106–3119.
- (21) Vosch, T., Cotlet, M., Hofkens, J., Van Der Biest, K., Lor, M., Weston, K., Tinnefeld, P., Sauer, M., Latterini, L., Müllen, K., and De Schrijver, F. C. (2003) Probing Förster type energy pathways in a first generation rigid dendrimer bearing two perylene imide chromophores. *J. Phys. Chem. A* 107, 6920–6931.
- (22) Förster, V. T. (1949) Experimentelle und theoretische Untersuchung des zwischenmolekularen Übergangs von Elektronenanregungsenergie. *Z. Naturforsch.* 4A, 321–327.
- (23) Ahn, K.-S., Ou, W., and Silver, J. (2004) Inhibition of certain strains of HIV-1 by cell surface polyanions in the form of cholesterol-labeled oligonucleotides. *Virology* 330, 50–61.
- (24) Boutorine, A. S., and Kostina, E. V. (1993) Reversible covalent attachment of cholesterol to oligodeoxyribonucleotides for studies of the mechanisms of their penetration into eucaryotic cells. *Biochimie* 75, 35–41.

BC050022G